

**INTENDED USE**

The COVID-19 & Influenza A/B & Adenovirus & RSV Antigen Nasal Test Kit is an *in vitro* immunoassay. The assay is for the direct and qualitative detection of antigens of SARS-CoV-2 (COV), Influenza A virus (FLU A), Influenza B virus (FLU B) adenovirus (ADV), and respiratory syncytial virus (RSV) from nasal secretions. The test is for professional use. Negative results do not preclude these viral infections. Testing results should not be the sole basis for treatment or other management decisions.

**PRINCIPLE**

The COVID-19 & Influenza A/B & Adenovirus & RSV Antigen Nasal Test Kit detects viral antigens through visual interpretation of color development on the three internal test strips for COV, FLU A/B and RAC (ADV+RSV combo) respectively.

**For COVID-19 test:**

Anti-SARS-CoV-2 antibodies are immobilized at the test region of the nitrocellulose membrane. Anti-SARS-CoV-2 antibodies conjugated to colored particles are immobilized on the conjugated pad. The nasal secretions, collected by the intended user, are supposed to be mixed with the extraction buffer, which is individually packed in the kit. During testing, target antigens, if present in the nasal secretions, will be released into the extraction buffer. As the specimen migrates along the strip by capillary action and then interacts with reagents on the Sample Pad, the target antigens will bind to anti-SARS-CoV-2 antibodies on the Conjugate Pad. Consequently, the antigen-antibody complex will be captured by the anti-SARS-CoV-2 antibodies immobilized at the Test Region. Excess colored particles will be captured at the Control Region of the NC membrane.

The presence of a colored band in the test region indicates a positive result for the SARS-CoV-2 viral antigens, while its absence indicates a negative result. A colored band at the control region serves as a procedural control, generally indicating that a proper volume of specimen has been added and membrane wicking is working.

**For Influenza A/B test:**

Anti-Influenza A virus antibodies and anti-Influenza B virus antibodies are immobilized at two separate test regions of the nitrocellulose membrane. Anti-Influenza A virus antibodies and anti-Influenza B virus antibodies conjugated to colored particles are immobilized on the conjugated pad. The nasal secretions, collected by the intended user, are supposed to be mixed with the extraction buffer, which is individually packed in the kit.

During testing, target antigens, if present in the nasal secretions, will be released into the extraction buffer. As the specimen migrates along the strip by capillary action and then interacts with reagents on the Sample Pad, the target antigens will bind to antibodies on the Conjugate Pad. Consequently, the antigen-antibody complex will be captured by the antibodies immobilized at the two Test Regions. Excess colored particles will be captured at the Control Region of the NC membrane.

The presence of a colored band in the test region indicates a positive result for the Influenza A/B viral antigens, while its absence indicates a negative result. A colored band at the control region serves as a procedural control, generally indicating that a proper volume of specimen has been added and membrane wicking is working.

**For RAC (ADV+RSV) test:**

Anti-adenovirus antibodies and Anti-respiratory syncytial virus antibodies are immobilized at the test region of the nitrocellulose membrane. Anti-adenovirus antibodies and Anti-respiratory syncytial virus antibodies conjugated to colored particles are immobilized on the conjugated pad.

The nasal secretions, collected by the intended user, are supposed to be mixed with the extraction buffer, which is individually packed in the kit.

During testing, target antigens, if present in the nasal secretions, will be released into the extraction buffer. As the specimen migrates along the strip by capillary action and then interacts with reagents on the Sample Pad, the target antigens will bind to anti-adenovirus antibodies or anti-respiratory syncytial virus antibodies on the Conjugate Pad. Consequently, the antigen-antibody complex will be captured by the anti-adenovirus antibodies or anti-respiratory syncytial virus antibodies immobilized at the Test Region. Excess colored particles will be captured at the Control Region of the NC membrane.

The presence of a colored band in the test region indicates a positive result for the adenovirus antigens or respiratory syncytial virus antigens, while its absence indicates a negative result. A colored band at the control region serves as a procedural control, generally indicating that a proper volume of specimen has been added and membrane wicking is working.

**MATERIALS**

**Materials Provided**

- Individually packed test
- Package insert

**Materials Required but Not provided**

- Clock, timer or stopwatch

**PRECAUTIONS**

- For *in vitro* Diagnostic Use Only.
- Caution should be taken when inserting the sample collector into the nasal cavity.
- DO NOT ingest.
- Read the *Package Insert* prior to use. Directions should be read and followed carefully.
- Do not use kit or components beyond the expiration date.
- The device contains material of animal origin and should be handled as a potential biohazard. Do not use if pouch is damaged or open.
- Test devices are packaged in foil pouches that exclude moisture during storage. Inspect each foil pouch before opening. Do not use devices that have holes in the foil or where the pouch has not been completely sealed. Erroneous result may occur if test reagents or components are improperly stored.
- All patient specimens should be handled and discarded as if they are biologically hazardous. All specimens must be mixed thoroughly before testing to ensure a representative sample prior to testing.
- Failure to bring specimens and reagents to room temperature before testing may decrease assay sensitivity. Inaccurate or inappropriate specimen collection, storage, and transport may yield false negative test results.
- Avoid skin or eyes contact with buffer before, during or after testing.
- Do not puncture the sealing membrane in the extraction tube before testing.

**STORAGE AND STABILITY**

- Store The COVID-19 & Influenza A/B & Adenovirus & RSV Antigen Nasal Test Kit at 2-30 °C when not in use.
- **DO NOT FREEZE.**
- Kit contents are stable until the expiration dates marked on their outer packaging and containers.

**TEST PROCEDURE**

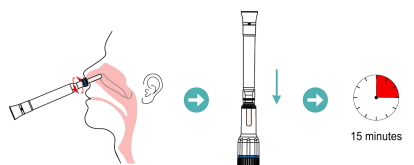
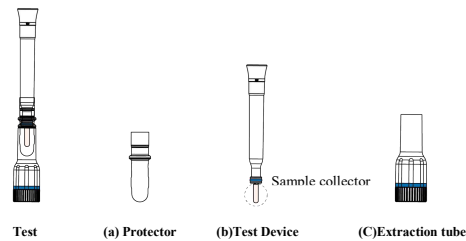
Bring devices, reagents and specimens and/or controls to room temperature (15-30°C) before use.

1. Remove the test from its packing. Label the device with the patient's identification. For best results, the assay should be performed within one hour.
2. 1) Take the test device out of the extraction tube.  
2) Remove the protector.
3. Gently insert the sample collector (the circle part in the picture) until resistance is met (about 1-2 cm into the nostril).
4. Rotate the collector **five times** against the nasal wall and remove from the nostril.
5. **Repeat the sample collection procedure for the other nostril** to ensure that sufficient specimen be collected from both nasal cavities.

**Note: 1. It is important to obtain as much secretion as possible.**

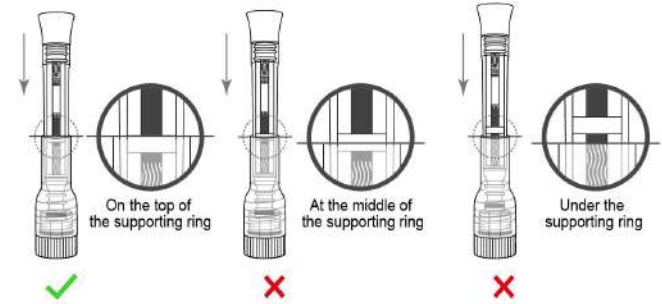
**2. This may feel uncomfortable. Do not insert the collector any deeper if patients feel strong resistance.**

6. Place the test device vertically into the extraction tube until the top edge of the extraction tube reach the top of the supporting ring.
7. Read the results at 15 minutes.



**Note:**

When placing the test device vertically into the extraction tube, the edge of the extraction tube must reach the top of the supporting ring. If not, this may lead to lateral flow failure, resulting in an incorrect result or invalid result.

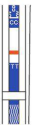


**RESULT INTERPRETATION**

**For COVID-19 test:**

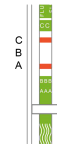


**POSITIVE: Two colored bands appear on the membrane.** One band appears in the control region (C) and another band appears in the test region (T).

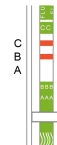


**NEGATIVE: Only one colored band appears, in the control region (C).** No apparent colored band appears in the test region (T).

**For Influenza A/B test:**



**Influenza A Positive:** One colored band appears in the control region (C), and another colored band in the A region (A).



**Influenza B Positive:** One colored band appears in the control region (C), and another colored band in the B region (B).



**Influenza A+B Positive:** One colored band appears in the control region (C), and two other colored bands appear in both A region (A) and B region (B).

**NOTE:** Co-infection with influenza A and B is rare. A clinical specimen that generates positive results for both A and B should be considered an invalid result, and another test should be performed. If the test is again positive for both influenza A and B, the specimen should be re-tested by another method prior to reporting of results.

**Negative:** Only one colored band appears in the control region (C), and band appears neither in the A region (A) nor B region (B).

**Invalid:** No colored band appears in the control region (C), whether a test band(s) is present or not. Repeat invalid tests with a new sample, new test device and reagent. Insufficient sample volume, inaccurate operating procedure or expired tests may yield an invalid result. Contact your local distributor if the problem continues.

**For RAC test:**

**RSV & ADV Positive:** One colored band appears in the control region (C), and two other colored bands appear in both R region (R) and A region (A).

**RSV Positive:** One colored band appears in the control region (C), and another colored bands appear in R region (R).

**ADV Positive:** One colored band appears in the control region (C), and another colored bands appear in A region (A).

**NEGATIVE:** Only one colored band appears, in the control region (C). No apparent colored band appears in the test region (R/A).

**INVALID:** Control band fails to appear. Results from any test which has not produced a control band at the specified read time must be discarded. Please review the procedure and repeat with a new test. If the problem persists, discontinue using the kit immediately and contact your local distributor.

**NOTE:**

- The color intensity in the test region(s) may vary depending on the concentration of analytes present in the specimen. Note that this is a qualitative test only, and cannot determine the concentration of analytes in the specimen.

**QUALITY CONTROL**

**Internal Procedural Controls**

The COVID-19 & Influenza A/B& Adenovirus&RSV Antigen Nasal Test Kit has built-in (procedural) controls. Each test has an internal standard zone to ensure proper sample flow. The user should confirm that the colored band located at the "C" region is present before reading the result.

**External Positive and Negative Controls**

Good laboratory practice suggests testing positive and negative external controls to ensure that the test reagents are working and that the test is correctly performed.

**LIMITATIONS OF THE TEST**

- The COVID-19 & Influenza A/B& Adenovirus&RSV Antigen Nasal Test Kit is for professional *in vitro* diagnostic use, and should only be used for the qualitative detection of antigens specific for SARS-CoV-2, Influenza A virus, Influenza B virus adenovirus, and respiratory syncytial virus. The intensity of color in a positive band should not be evaluated as "quantitative or semi-quantitative".

- Both viable and nonviable viruses are detectable with the kit.
- As with all diagnostic tests, a definitive clinical diagnosis should not be based on the results of a single test, but should only be made by the physician after all clinical and laboratory findings have been evaluated.
- Failure to follow the TEST PROCEDURE and RESULT INTERPRETATION may adversely affect test performance and/or invalidate the test result.
- Results obtained with this assay, particularly in the case of weak test lines that are difficult to interpret, should be used in conjunction with other clinical information available to the physician.
- Negative results do not preclude viral infections and should be confirmed via molecular assay.

**PERFORMANCE CHARACTERISTICS**

**Analytical Sensitivity:**

Items	LOD
SARS-CoV-2	1 × 10 <sup>2</sup> .4 TCID50/mL
Influenza A virus (H1N1)	1 × 10 <sup>4</sup> TCID50/mL
Influenza A virus (H3N2)	3.125 × 10 <sup>3</sup> TCID50/mL
Influenza B virus (Yamagata)	2.5 × 10 <sup>4</sup> TCID50/mL
Influenza B virus (Victoria)	6.5 × 10 <sup>3</sup> TCID50/mL
Respiratory syncytial virus (Type A)	2.4 × 10 <sup>4</sup> TCID50/mL
Respiratory syncytial virus (Type B)	2.5 × 10 <sup>3</sup> TCID50/mL
Adenovirus	2.4 × 10 <sup>4</sup> TCID50/mL

**Clinical Evaluation:**

**For COVID-19 Test:**

A total of 151 clinical specimens were collected to verify the performance of COVID-19 Antigen Test. Individuals who are suspected of COVID-19 were enrolled in this study. 32 positive specimens and 119 negative specimens were confirmed by RT-PCR.

Table 1: Clinical Summary of COVID-19 Antigen

		RT-PCR		Total
		Positive	Negative	
COVID-19 Antigen Test	Positive	31	0	31
	Negative	1	119	120
	Total	32	119	151

Relative Sensitivity: 96.9% (84.3%–99.4%)\*  
 Relative Specificity: 99.9% (96.9%–100.0%)\*  
 Overall Agreement: 99.3% (96.3%–99.9%)\*  
 \*95% Confidence Interval

**For FLU A/B Test:**

For Influenza A Test:

A total of 151 clinical specimens were collected, to verify the performance of Influenza A/B Antigen Test. 25 were found to be positive by RT-PCR and 126 were found to be negative by RT-PCR.

For Influenza B Test:

A total of 151 clinical specimens were collected, to verify the performance of Influenza A/B Antigen Test. 25 were found to be positive by RT-PCR and 126 were found to be negative by RT-PCR.

Table 2: Clinical Summary of Influenza A

		RT-PCR		Total
		Positive	Negative	
Influenza A Test	Positive	25	0	25
	Negative	0	126	126
	Total	25	126	151

Relative Sensitivity: 99.9% (86.7%–100.0%)\*  
 Relative Specificity: 99.9% (97.0%–100.0%)\*  
 Overall Agreement: 99.9% (97.5%–100.0%)\*  
 \*95% Confidence Interval

Table 3: Clinical Summary of Influenza B

		RT-PCR		Total
		Positive	Negative	
Influenza B	Positive	25	0	25

Test	Negative	1	125	126
	Total	26	125	151

Relative Sensitivity: 96.2% (81.1%–99.3%)\*  
 Relative Specificity: 99.9% (97.0%–100.0%)\*  
 Overall Agreement: 99.3% (96.3%–99.9%)\*  
 \*95% Confidence Interval

**For Adenovirus (ADV) test:**

A total of 151 clinical specimens were collected, to verify the performance of Adenovirus Antigen Test. 18 were found to be positive by RT-PCR and 133 were found to be negative by RT-PCR.

Table 1: Clinical Summary of ADV

		RT-PCR		Total
		Positive	Negative	
Adenovirus Test	Positive	18	1	19
	Negative	0	132	132
	Total	18	133	151

Relative Sensitivity: 99.9% (82.4%–100.0%)\*  
 Relative Specificity: 99.2% (95.9%–99.9%)\*  
 Overall Agreement: 99.3% (96.3%–99.9%)\*  
 \*95% Confidence Interval

**For Respiratory Syncytial Virus (RSV) test:**

A total of 151 clinical specimens were collected, to verify the performance of Respiratory Syncytial Virus Antigen Test. 18 were found to be positive by RT-PCR and 133 were found to be negative by RT-PCR.

Table 4: Clinical Summary of RSV

		RT-PCR		Total
		Positive	Negative	
Respiratory Syncytial Virus Test	Positive	18	1	19
	Negative	0	132	132
	Total	18	133	151

Relative Sensitivity: 99.9% (82.4%–100.0%)\*  
 Relative Specificity: 99.2% (95.9%–99.9%)\*  
 Overall Agreement: 99.3% (96.3%–99.9%)\*  
 \*95% Confidence Interval

**Cross Reactivity:**

Cross reactivity with the following organisms has been studied. Samples positive for the following organisms were found negative when tested with The COVID-19 & Influenza A/B& Adenovirus&RSV Antigen Nasal Test Kit.

Epstein-Barr virus	Parainfluenza virus	Haemophilus influenzae
Enterovirus EV70	Parainfluenza virus	Legionella pneumophila
Enterovirus EV71	Parainfluenza virus	Mycoplasma pneumoniae
Enterovirus A16	Parainfluenza virus	Mycobacterium tuberculosis
Enterovirus A24	Respiratory syncytial virus A	Staphylococcus aureus
Enterovirus B1	Respiratory syncytial virus B	Staphylococcus epidermidis
Echovirus 6	Rhinovirus A30	Streptococcus agalactiae
HCoV-229E	Rhinovirus B52	Streptococcus pneumoniae
HCoV-OC43	Bordetella parapertussis	Streptococcus pyogenes
HCoV-NL63	Bordetella pertussis	
MERS-coronavirus	Candida albicans	
Human metapneumovirus	Chlamydia pneumoniae	
Norovirus	Group C Streptococcus	

**NOTE:**

- For COV test:** COVID-19 detection has no across reactivity with influenza A, influenza B and respiratory syncytial virus and adenovirus.
- For ADV test:** Adenovirus detection has no across reactivity with COVID-19, influenza A, influenza B and respiratory syncytial virus.
- For FLU A/B test:** FLUA detection has no across reactivity with influenza B, respiratory syncytial virus, adenovirus. FLUB detection has no across reactivity with COVID-19, influenza A, respiratory syncytial virus, adenovirus.
- For RSV test:** RSV detection has no across reactivity with COVID-19, influenza A, influenza B

and adenovirus.

**Interfering Substances**












The following substances, naturally present in respiratory specimens or that may be artificially introduced into the respiratory tract, were evaluated at the concentrations listed below. None of them were found to affect test performance of the kit.

Substance	Concentration	Substance	Concentration
3 OTC nasal sprays	10%	Guaiacol glyceryl ether	20mg/mL
3 OTC mouth washes	10%	Mucin	1%
3 OTC throat drops	10%	Whole blood	4%
4-acetamidophenol	10 mg/mL	Mupirocin	250µg/mL
Acetylsalicylic acid	10 mg/mL	Oxymetazoline	25µg/mL
Albuterol	10 mg/mL	Phenylephrine	10 mg/mL
Chlorpheniramine	5 mg/mL	Phenylpropanolamine	1mg/mL
Dexamethasone	50µg/mL	Zanamivir	10mg/mL
Dextromethorphan	10µg/mL	Adamantanamine	500 ng/mL
Diphenhydramine	5 mg/mL	Oseltamivir phosphate	10mg/mL
Doxylamine succinate	1 mg/mL	Tobramycin	10mg/mL
Flunisolide	25µg/mL	Triamcinolone	14mg/mL

**LITERATURE REFERENCES**

- Voss, Jameson D.; Atkinson, Richard L.; Dhurandhar, Nikhil V. (1 November 2015). "Role of adenoviruses in obesity". *Rev. Med. Virol.* 25(6): 379–387. doi:10.1002/rmv.1852. PMID 26352001.
- Wadell G.; et al. (1987). Whelan, Julie; Bock, Gregory (eds.). *Novel diarrhoea viruses*. New York: Wiley. p. 63. ISBN 978-0-471-91094-7.
- Wikipedi acontributors, "Influenza virus C," Wikipedia, The Free Encyclopedia,http://en.wikipedia.org/w/index.php?title=Influenzavirus\_C&oldid=649896527 (accessed May 25, 2015).
- Course BS3035: Virology, University of Leicester, http://www-micro.msb.le.ac.uk/3035/Paramyxoviruses.html.
- Respiratory Syncytial Virus (RSV): Overview, Treatment, and Prevention Strategies, Mark J. Polak, MD.
- Macartney K. et al. Nosocomial Respiratory Syncytial Virus Infections: The Cost-Effectiveness and Cost-Benefit of Infection Control. *Pediatrics* Vol. 106 No. 3 Sept 2000, pp. 520–526. http://pediatrics.aappublications.org/cgi/content/full/106/3/520.
- Thompson W. et al. Mortality Associated With Influenza and Respiratory Syncytial Virus in the United States. *JAMA*, January 8, 2003 – Vol 289, No. 2.
- Moler F.W. et al. Respiratory syncytial virus morbidity and mortality estimates in congenital heart disease patients: a recent experience. *Crit Care Med.* 1992 Oct; 20(10):1406–13.

**GLOSSARY OF SYMBOLS**

	Catalog number		Temperature limitation
	Consult instructions for use		Batch code
	In vitro diagnostic medical device		Use by
	Manufacturer		Contains sufficient for <n> tests
	Do not reuse		Authorized representative in the European Community
	CE marking according to IVD Medical Devices Directive 98/79/EC		



Assure Tech. (Hangzhou) Co., Ltd.  
 Building 4, No. 1418-50, Moganshan Road,  
 Gongshu District, Hangzhou,  
 310011 Zhejiang, P.R. China  
 contact@diareagent.com



MedUnion S.L.  
 Carrer de Tapioles,33, 2-1, Barcelona, 08004,  
 Spain  
 E-mail: admin@medunion.es